

Enhanced β -Carotene Biosynthesis in Recombinant *Escherichia coli* Harboring the Bottom Portion of Mevalonate Pathway of *Enterococcus faecium* VTCC-B-935 Isolated in Vietnam

Abstract

β -carotene is a carotenoid pigment which has commercial value. Biosynthesis of this pigment using recombinant *E. coli* has been reported. However, improvement of yield is remained necessary. In this case report, bottom portion of mevalonate partway was employed for enhancing metabolic flow to isopentenyl diphosphate, the building block of all carotenoid. Three genes including *mvaK1*, *mvaK2*, and *mvaD* were cloned from *Enterococcus faecium* VTCC-B-935 isolated in Vietnam and placed into pET28a(+) vector resulted in pET28-K1K2D. Co-expression of this vector with pRSET-IEIBY which contains five enzymes responsible for biosynthesis of β -carotene in *E. coli* BL21 (DE3) resulted in approximately three fold higher yield of the compound. The high copy number vector pRSET-A showed better performance in production of β -carotene over low copy pET-22b(+). Addition of glycerol significantly enhanced *E. coli* cell growth and β -carotene biosynthesis.

Keywords: β -carotene; *Enterococcus faecium* VTCC-B-935; *mvaK1*; *mvaK2*; *mvaD*; Mevalonate pathway

Research Article

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Abbreviations: IPP: Isopentenyl Diphosphate; MEP: 2-C-Methyl-D-Erythritol-4-Phosphate; MVA: Mevalonate; DMAPP: Dimethylallyl Diphosphate; FPP: Farnesyl Diphosphate; GGPP: Geranylgeranyl Diphosphate; G3P: Glyceraldehyde-3-Phosphate

Introduction

β -carotene, one of the carotenoid compounds, was found mainly in plants. Because of its highly antioxidant activity, this compound was shown to have good effects to human health. It is the precursor of vitamin A which is critically required for human [1]. Due to good health effects and yellow color, β -carotene is widely used in industries including nutraceuticals, pharmaceuticals, food colorants, cosmetics, and animal feed additives. However, production of natural β -carotene is not sufficient to afford demand of the market. Currently, more than 90% of commercially available β -carotene is chemically synthesized [2]. This fact leads a number of research groups to the trend of seeking for alternative sources of natural β -carotene. Natural carotenogenic microorganisms, e.g. *Blakeslea trispora*, *Rhodotorula glutinis*, and *Dunaliella salina* have been used for fermentation to produce β -carotene [3-5].

The availability of carotenoid genes from natural carotenogenic organisms leads to another strategy of natural carotenoid biosynthesis. Carotenoid genes were cloned and introduced into non-carotenogenic organism, *Escherichia coli*, for production of carotenoid, including β -carotene [6-19]. *E. coli* is considered as one of the ideal hosts because of its convenience genetic engineering system and fast growth. However, to date, the production of

β -carotene is still unable to support industrial needs because of insufficient yield and stability. Therefore, attempts are required for improvement of production of carotenoid in general and β -carotene in particular. Biosynthesis of β -carotene was affected by a number of factors. In our study, we focus in

- i. The different expression vector backbones
- ii. Origin of foreign genes
- iii. Availability and balance of isopentenyl diphosphate (IPP) which is the building blocks of carotenoids, and
- iv. Effect of additional carbon sources

Biosynthesis partway of β -carotene is indicated in Figure 1. DMAPP and IPP, the building blocks of carotenoid are synthesized via 2-C-methyl-d-erythritol-4-phosphate (MEP) pathway which is autonomous in our chosen host *E. coli*. However, in theory the natural yield of these precursors is only sufficient for natural need of *E. coli* in normal growth conditions which is far less than that required once the organism is used as a microbial factory to produce β -carotene. To address this issue, addition of the exogenous mevalonate (MVA) pathway was shown as a reasonable strategy [6,12,18,20,21]. The MVA pathway is divided into two portions, the upper (from acetyl-CoA to MVA) and the lower (from MVA to DMAPP and IPP).

Natural *E. coli* harbors MEP pathway that enable biosynthesis of FPP from G3P and pyruvate, as well as IPP isomerase catalyze the two-way conversion of IPP and DMAPP, the precursors

of carotenoid (yellow part). These precursors could also be synthesized by the mevalonate pathway which is exogenous to *E. coli*. In this design, the bottom portion of MVA pathway from mevalonate to IPP was recruited from *E. faecium* VTCC-B-935 isolated in Vietnam (green part). IPP is subsequently used as building blocks for the process of β -carotene synthesis including

four steps catalyzed by exogenous *crt* genes derived from *P. ananatis*. For better balancing between the two isoforms IPP and DMAPP, in addition to the endogenous *idi* gene positioned in the genome of *E. coli* host, another copy of this gene was introduced into the carotenogenic vector for co-over expression with *crt* genes (blue part).

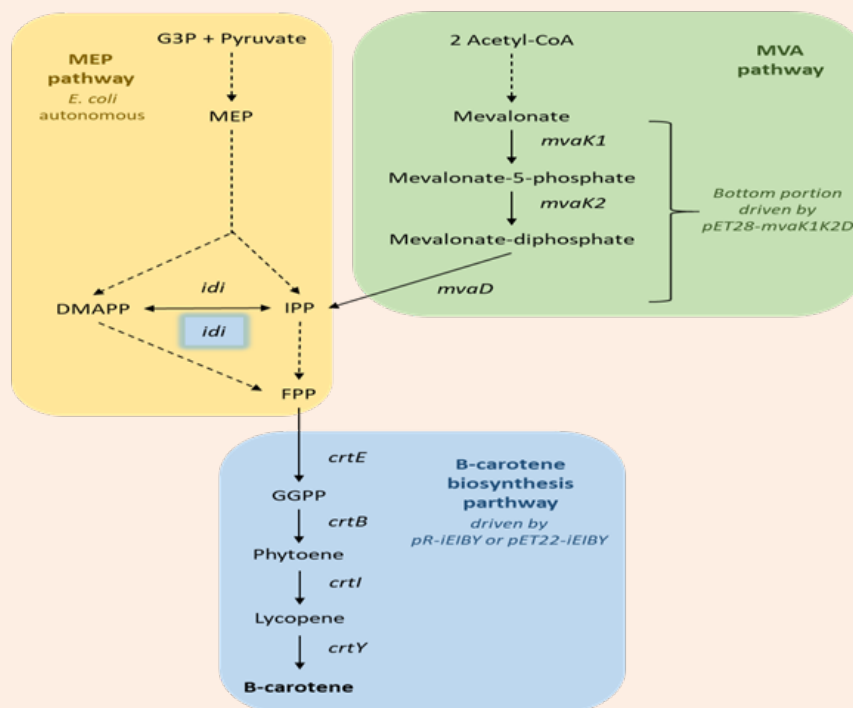


Figure 1: Biosynthesis pathways of β -carotene in recombinant *E. coli* engineered in this study.

G3P: Glyceraldehyde-3-Phosphate; MEP: 2-C-Methyl-d-Erythritol-4-Phosphate; IPP: Isopentenyl Diphosphate; DMAPP: Dimethylallyl Diphosphate; FPP: Farnesyl Diphosphate; GGPP: Geranylgeranyl Diphosphate

In this case report, we first examine the difference of expression vector backbones. In our previous study, three genes including *crtE*, *crtB*, and *crtI* were cloned from *Pantoea ananatis* for lycopene biosynthesis, and *idi* was cloned from *E. coli* for better balancing of IPP [7]. These four genes, together with *crtY*, were introduced into pRSET-A and pET22b(+) resulted in multicistronic operon vectors pR-IEIBY and pET22-IEIBY, respectively. Secondly, differ from other previous publications, another source of genes encode for the bottom mevalonate pathway enzymes including *mvaK1*, *mvaK2*, and *mvaD* were cloned from *Enterococcus faecium* VTCC-B-935 isolated in Vietnam and introduced into pET28 vector forming pET28-mvaK1K2D. Subsequently, this vector was transformed into a recombinant *E. coli* strain which has already contained another expression vector pR-IEIBY. Finally, four additional carbon sources were investigated for higher production of β -carotene using our recombinant system.

Materials and Methods

Bacterial strains, primers, plasmids, and culture conditions

Bacterial strains, primers, and plasmids used in this study are listed in Table 1. *E. coli* DH5 α was used for molecular cloning

of target genes. *E. coli* BL21(DE3) was used for expression of mevalonate and carotenoid genes and subsequently the expressed recombinant enzymes produced β -carotene. *Enterococcus faecium* VTCC-B-935 and *Pantoea ananatis* DSM-17873 were used as donors of target genes. Primer sequences were designed based on nucleotide sequences of target genes available in NCBI database, D90087.2 for *crtY* and AF290095.1 for *mva* genes. Backbone plasmids including pLUG, pRSET-A, pET22b(+) and pET28a(+) were used for gene cloning and recombinant enzyme expression.

Cultures of *E. faecium* and *P. ananatis* were performed following provider's instructions. For β -carotene production, recombinant *E. coli* clones were grown on LB agar plates under selection pressure of ampicillin (100 μ g/mL) or combination of ampicillin/kanamycin (50 μ g/mL) in an incubator at 37°C overnight. Individual colonies on plates were inoculated in a flask containing 5 ml of LB media supplemented with the same antibiotic for 12h in a rotary-shaking incubator at 180 RPM. Subsequently the primary culture was transferred into 250mL flask containing 50mL LB media under the same condition. IPTG induction, at different concentrations, was performed while OD600nm reached 0.3-0.5 and continue inoculation for desired periods. Conditions were modified in optimization experiments

according to planned changes. For biosynthesis of IPP and DMAPP by bottom portion of MVA pathway, mevalonate was added to the culture at 6.6mM as substrate [13].

Cloning of crtY, mvaK1, mvaK2, and mvaD genes

Genomic DNA of *P. ananatis* and *E. faecium* were extracted using common protocols. Target genes were amplified using standard protocols and specific primers listed in Table 1. PCR products were subsequently ligated into pLUG cloning vector (Intron Biotechnology) by T4 DNA ligase (New England Biolabs).

Ligation products were transformed into competent *E. coli* DH5 α (Thermo Fisher Scientific) cells by heat shock and plated onto LB agar plates supplemented with ampicillin (100 μ g/mL), X-gal (20 μ g/mL) and IPTG (1 mM). White colonies were picked and inoculated in 5 ml LB media. After extraction, plasmids were screened by size separation by agarose electrophoresis, and then mapped by appropriate restriction enzymes. Target inserts in positive clones were sequenced by Marcogen. Gene sequences were in silico analyzed by Vector NTI software to ensure correct translation, and aligned to available database using BLAST.

Table 1: Bacterial strains, primers, and plasmids used in this study.

Name	Description	Reference/Source
Bacteria Strains		
<i>E. coli</i> DH5 α	TA cloning host	Thermo Fisher Scientific
<i>E. coli</i> BL21(DE3)	Expression host compatible with vectors driven by T7 system	Thermo Fisher Scientific
<i>Pantoea ananatis</i> DSM-17873	Carotenoid genes donor	German Collection of Microorganisms and Cell Cultures
<i>Enterococcus faecium</i> VTCC-B-935	Mevalonate genes donor, isolated in Vietnam	Vietnam Type Culture Collection
Primers*		
mvaK1-F	ggatcc ATGGCAA ACTATGGCCAAG	This study
mvaK1-R	gaattc TAAAC ATAGGTATGTACTCCT	This study
mvaK2-F	gaattc ATGATTGA AGTATCTGCACCA	This study
mvaK2-R	gagctc TCATCGG TTTTCTCTTTGA	This study
mvaD-F	gagctc ATGTTTAA AGGCAAAGCACG	This study
mvaD-R	gcggccgc TTATTCA AATCGCAATTCCTG	This study
crtY-F	TgaattcAGGAGGTGCTCT TAAAT GGGAGCGGCTAT	This study
crtY-R	GCaagctt TAA CGATGAGTCGTCATAATGG	This study
Plasmids		
pLUG	TA cloning vector	Intron Biotechnology
pET28a(+)	Expression vector for low copy number (pBR322 ori), strong phage promoter (T7), IPTG induction (lac operon), and Kanamycin selection (Kan ^r)	Novagen
pET22b(+)	Expression vector for low copy number (pBR322 ori), periplasmic localization (pelB signal peptide), strong phage promoter (T7), IPTG induction (lac operon), and Ampicillin selection (Amp ^r)	Novagen
pRSET-A	Expression vector for high copy number (pUC ori), strong phage promoter (T7), IPTG induction (lac operon), and Ampicillin selection (Amp ^r)	Thermo Fisher Scientific
pR-iEIB	pRSET-A containing idi of <i>E. coli</i> and crtE, crtI and crtB of <i>P. ananatis</i>	[7]
pR-iEIBY	pRSET-A containing idi of <i>E. coli</i> and crtE, crtI, crtB, and crtY of <i>P. ananatis</i>	This study
pET22-Y	pET22b(+) containing crtY of <i>P. ananatis</i>	This study
pET22-iEIBY	pET22 containing idi of <i>E. coli</i> and crtE, crtI, crtB, and crtY of <i>P. ananatis</i>	This study
pET28-K1	pET28a(+) containing mvaK1 of <i>E. faecium</i>	This study
pET28-K1K2	pET28a(+) containing mvaK1 and mvaK2 of <i>E. faecium</i>	This study
pET28-K1K2D	pET28a(+) containing mvaK1, mvaK2, and mvaD of <i>E. faecium</i>	This study

(*) Template binding sequences are indicated by underlined letters, start and stop codons are bold, and restriction sites are lowercase. In cases of no restriction sites overhanged, autonomous sites in the carrying vectors were used instead.

Construction of recombinant plasmids

In our previous study, four genes including idi, crtE, crtI, and

crtB were introduced into pRSET-A resulted in pR-IEIB which was used for biosynthesis of lycopene [7]. In this study, cloned crtY gene was excised from pLUG-crtY and inserted into pR-IEIB using

HindIII restriction enzyme resulted in pR-IEIBY. Subsequently, the multicistronic operon in pR-IEIBY containing five genes was moved to pET22b(+) by two steps. First, only *crtY* was moved using EcoRI and HindIII resulted in pET22-Y. Second, the fragment containing four genes *idi*, *crtE*, *crtI* and *crtB* was moved to pET22-Y using XbaI and EcoRI resulted in pET22-IEIBY. Similar

experimental procedure was performed to introduce *mvaK1*, *mvaK2*, and *mvaD* into pET28a(+) to construct pET28-K1, pET28-K1K2, and pET28-K1K1D at the positions of BamHI/EcoRI, EcoRI/SacI, and SacI/NotI, respectively. All vectors constructed in this study are listed in Table 1 and the maps of three major vectors are presented in Figure 2.

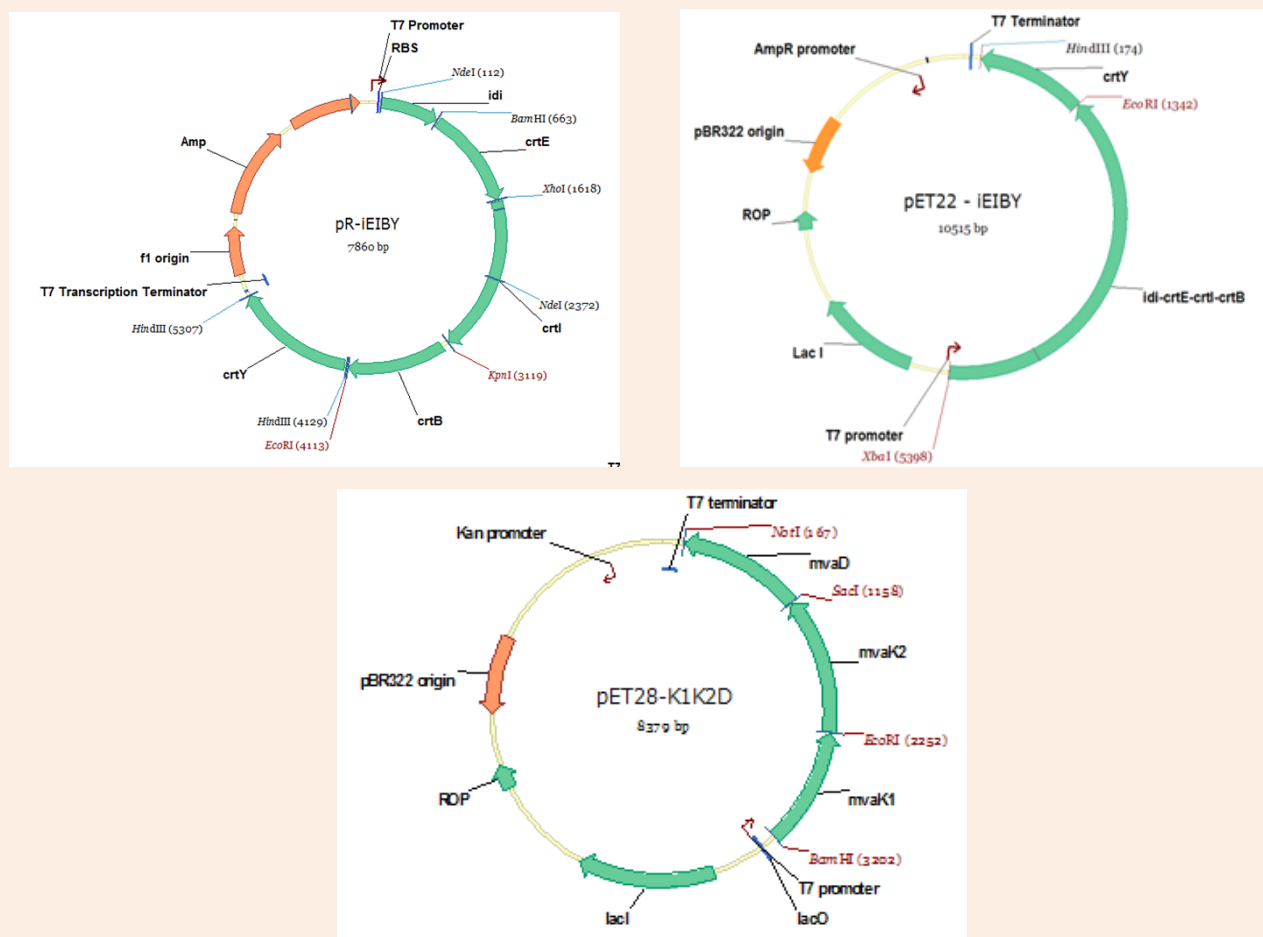


Figure 2: Expression vectors used for biosynthesis of β -carotene in this study.

Where, pR-IEIBY and pET22-IEIBY are pRSET-A and pET22b(+) containing multicistronic operon of five genes including *idi* from *E. coli* DH5 α , *crtE*, *crtI*, *crtB*, and *crtY* from *Pantoea ananatis*, respectively. These two vector were used for biosynthesis of β -carotene in recombinant *E. coli* BL21(DE3). pET28-K1K2D is pET28a(+) containing multicistronic operon of three genes including *mvaK1*, *mvaK2*, and *mvaD* from *Enterococcus faecium* VTCC-B-935. This vector was used for conversion of mevalonate to IPP. Major component of the vectors and restriction sites used in the construction were indicated. Vector maps were drawn using Vector NTI software version 11.5.1.

Measurement of β -carotene production

Concentration of β -carotene synthesized by recombinant *E.*

coli was measured by HPLC according to the protocol previously described by Yoon et al. [12] with slight modifications. *E. coli* cells were harvested by centrifugation at 12,000 rpm and washed once by ddH₂O. Cell pellet was re-suspended in 1mL of acetone and incubate at 55°C for 10 to 15 minutes in dark condition. The extracted acetone containing β -carotene was collected by centrifugation at 12,000 rpm for 15 minutes. HPLC analysis was performed on the Agilent 1260 system using Zorbax C18 column (250 mm $\times$ 4.6 mm, 5 micromet) with detector at 454 nm. Two mobile phases were used including methanol (phase A) and acetonitrile (phase B). Isocratic program (A:B= 70:30) and flow rate of 1ml/min was used to all analyses. Standard β -carotene was purchased from Sigma (Cat.No. C4582).

Results and Discussion

Comparison of pRSET-A and pET22b(+) for biosynthesis of β -carotene in engineered *E. coli*

Different expression vectors produce different levels of desired recombinant proteins; in this case the five enzymes involved in the biosynthesis of β -carotene, and subsequently affect production of this carotenoid pigment. Therefore, we first compare two of the most common commercially available expression systems including pRSET-A and pET22b(+). Although these two vector systems are both driven by T7 promoter, they are significantly different in term of copy number. In case of carotenoid production by recombinant *E. coli*, contradictory observations regarding the biosynthesis efficiency caused by plasmid copy number were reported. Jones et al. [22] showed equal or better performance of low copy plasmids in comparison to high copy plasmids [22]. However, using a mutated version of the low copy vector pDC318 that made it a high copy one, Tao et al. [23] observed that mutated vector showed higher level of β -carotene biosynthesis [23]. In our study, accumulation of β -carotene was higher with pRSET-A than pET22b(+) according to higher density of yellow color of the cell pellet (Figure 3). HPLC analysis confirmed the biosynthesis of β -carotene by pR-IEIBY and pET22-IEIBY are 5.5mg/L and 3.1mg/L, respectively. pRSET-A harbor pUC origin of replication which manages approximately 500 copies of the vector per single cell. This number is about 10 to 20 folds higher than what driven by pBR322 origin of replication located in pET22b(+). Although pET22b(+) and pRSET-A are not totally identical but they are both driven by T7 promoter. Therefore, in term of plasmid copy number our data is in agreement with Tao et al. [23].

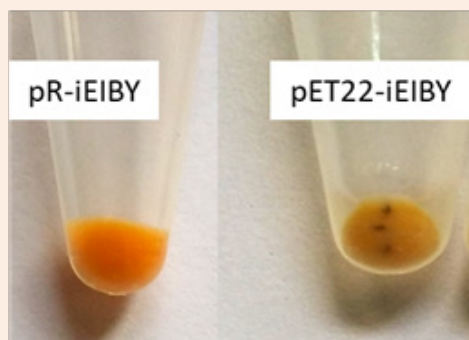


Figure 3: Comparison of pRSET-A and pET22b(+) for biosynthesis of β -carotene in recombinant *E. coli* BL21(DE3).

Higher accumulation of β -carotene in cell pellet of pR-IEIBY than in pET22-IEIBY was demonstrated by higher density of yellow color which is the color of this pigment.

Increased production of β -carotene by addition of bottom portion of mevalonate pathway

Isopentenyl diphosphate (IPP) and dimethylallyl diphosphate (DMAPP) are precursors of all carotenoid, including β -carotene. Therefore, to enhance production of β -carotene, it is reasonable to direct the biosynthesis flow of the *E. coli* host toward these building blocks via either endogenous MEP pathway or exogenous

MEV pathway, or both (Figure 1). Several studies have proven the feasibility of this strategy [6,12,13,18]. In genetic engineering studies focusing on production of recombinant proteins, it is common that different sources of target genes resulted in different level of expression because of the suitability between genetic code of foreign genes with the genetic reading system/mechanism of specific host. In this study, we test another set of bottom portion of MVA pathway derived from *E. faecium* isolated in Vietnam.

E. coli BL21 (DE3) harboring pR-IEIBY was made competent. This enabled second transformation with pET28-K1K2D resulted in a recombinant strain harboring both carotenoid vector and MVA vector, in this case are pR-IEIBY and pET28-K1K2D, respectively. *E. coli* culture was supplied with mevalonate at 6.6 mM which the optimal concentration is shown by Yoon et al. [13]. It is clear that addition of bottom portion of MVA pathway of *E. faecium* VTCC-B-935 did not affect growth of *E. coli* (Figure 4A). This observation is in agreement with several previous reports [6,11-13]. To date, β -carotene has not been shown as a toxic agent to *E. coli*. As expected, over expression of three *mva* genes significantly up regulates the biosynthesis of β -carotene (Figure 4B). At 48 hours and 96 hours of cultivation, addition of bottom MVA pathway derived from *E. faecium* VTCC-B-935 resulted in 12.6mg/L and 17.7mg/L of β -carotene which are approximately three folds higher than 4.2mg/L and 5.6mg/L, respectively, produced in control strain. This result indicated that the three genes, *mvaK1*, *mvaK2*, and *mvaD* of *E. faecium* VTCC-B-935 are comparable to the corresponding genes of *Streptococcus pneumoniae* in term of IPP biosynthesis in recombinant *E. coli* [13].

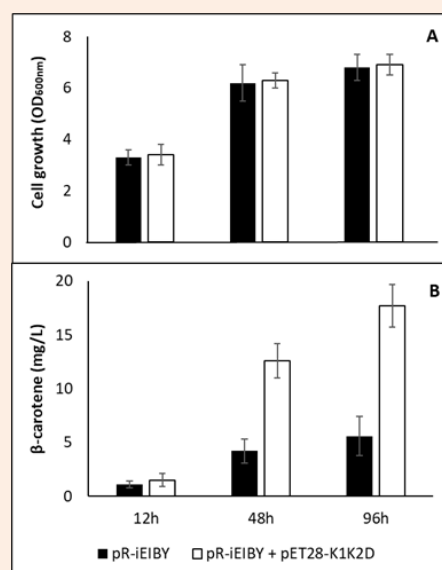


Figure 4: Cell growth and β -carotene production of recombinant *E. coli* with or without addition of bottom portion of MVA pathway from *E. faecium* VTCC-B-935.

pR-IEIBY contains β -carotene biosynthesis genes and *idi*. pET28-K1K2D contains genes encoding enzymes catalyze reactions from mevalonate to IPP.

Effect of addition of carbon sources on β -carotene production

Carbon source was determined as one of the factors related to biosynthesis of β -carotene. Insufficient availability of carbon sources restricts the growth of cell. Again, contradictory observations were reported regarding the effect of different carbon sources on biosynthesis of carotenoid by recombinant *E. coli*, including β -carotene. Biosynthesis of lycopene was shown increased and decreased following addition of glycerol and glucose, respectively [24]. Contradictorily, production of lycopene was significantly enhanced by addition of glucose in high cell density fed-batch fermentation conditions [25]. In this study, four sources of carbon were investigated, including glucose, galactose, maltose, and glycerol. The optimal concentration is 0.5% according to Yoon et al. [12]. Therefore, we added 0.5% of each of carbon sources to the 2YT medium. In Figure 5, the positive correlation between cell growth and β -carotene yield was observed. Among the tested carbon sources, glycerol indicated significantly improvement in both cell growth and β -carotene production over the others. In this case, the highest yield of β -carotene was 65 mg/L which was approximately three fold higher than the control which did not contain additional carbon source. Beside glycerol, addition of maltose slightly improved cell growth and β -carotene production. However, addition of glucose and galactose resulted in lower cell growth and β -carotene biosynthesis compared to the control. One of the reasons to explain this phenomenon is that glucose and galactose decreased pH of the culture that leads to inhibition of *E. coli* growth.

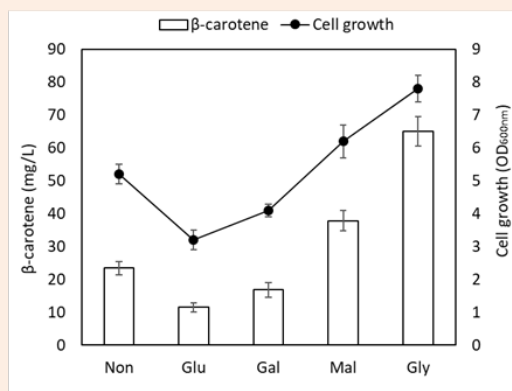


Figure 5: Effect of various carbon sources on growth and production of β -carotene by recombinant *E. coli* harboring pR-IEIBY and pET28-K1K2D.

Non: Control which did not contain any additional carbon source; Glu: Glucose; Gal: Galactose; Mal: Maltose; Gly: Glycerol

Conclusion

The bottom portion of mevalonate pathway of *E. faecium* VTCC-B-935 isolated in Vietnam was introduced into an engineered carotenogenic *E. coli* resulted in improvement of β -carotene biosynthesis. The pRSET-A expression vector performed better than pET22b(+). Addition of glycerol as carbon source enhanced

E. coli cell growth and biosynthesis of β -carotene almost 3 folds.

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